

# Convergent synthesis of (1→2)- and (1→4)-C-linked imino disaccharides†

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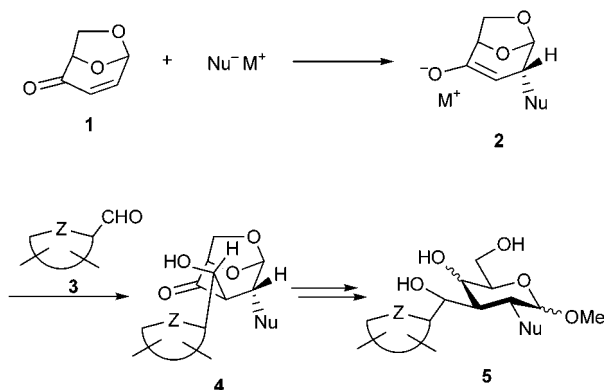
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**A convergent synthesis of (1→2)- and (1→4)-C-linked imino disaccharides was achieved by applying Nozaki–Kishi coupling of a hydroxyproline-derived carbaldehyde with isolevoglucosenone or levoglucosenone derived enol triflates.**

Carbohydrate mimetics are potentially useful tools to study cellular interactions,<sup>1</sup> the biosynthesis of glycoproteins, the metabolism of glycoconjugates,<sup>2</sup> and the mechanisms of digestion.<sup>3</sup> Inhibitors of the enzymes involved in these processes such as the glycosidases and the glycosyltransferases are potential anti-cancer, antiviral, and antidiabetic agents, as well as insect antifeedant agents.<sup>4</sup> Disaccharide mimetics such as C-disaccharides and dideoxyiminoalditol C-linked to monosaccharides have emerged as a new class of specific glycosidase inhibitors and may represent non-hydrolyzable epitopes.<sup>5–7</sup> Recently we disclosed an efficient and versatile approach to the syntheses of (1→3)-C-disaccharides and (1→3)-C-linked imino disaccharides **5** based on the cross-aldolisation of the aldehydes **3** with the nucleophile 1,4-adducts **2** of isolevoglucosenone **1** (Scheme 1).<sup>8,9</sup> We report here that (1→4)- and (1→2)-C-disaccharides can be prepared starting from **1** and levoglucosenone **17** with **3**, respectively.

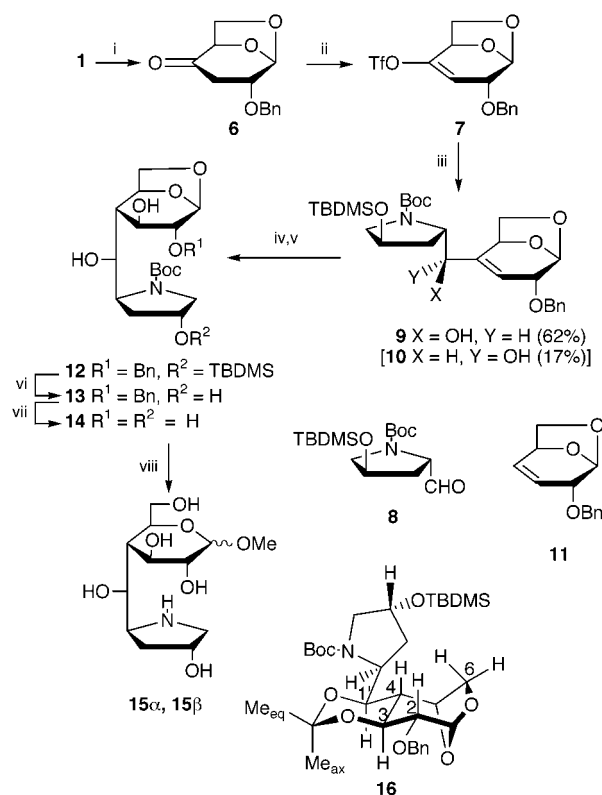
Adduct **6** of benzyl alcohol with isolevoglucosenone was enolized, without elimination of benzylate anion, on treatment with LiHMDS in 95:5 THF–HMPA at –78 °C (Scheme 2).<sup>9</sup> Quenching of the corresponding lithium enolate with 2-[bis(trifluoromethylsulfonyl)amino]-5-chloropyridine provided the enol triflate **7** in 87% yield.<sup>10</sup> Nozaki–Kishi coupling of **7** and aldehyde **8** led to allylic alcohols **9** and **10** isolated in 62 and 17% yield, respectively.<sup>11,12</sup> Interestingly, the reaction was accelerated by ultrasound and O<sub>2</sub>. Under N<sub>2</sub> atmosphere, ultrasound shortened the reaction time from 30 to 2 h. While in the presence of a catalytic amount of O<sub>2</sub> (5 mol% with respect to CrCl<sub>2</sub>, concentration of O<sub>2</sub> lower than 10%), the reaction time was reduced further to less than 1 h. Moreover, O<sub>2</sub> suppressed the formation of **11**, resulting from H<sub>2</sub>O quenching of the alkenylchromium species, from 36 to less than 10%.



Scheme 1

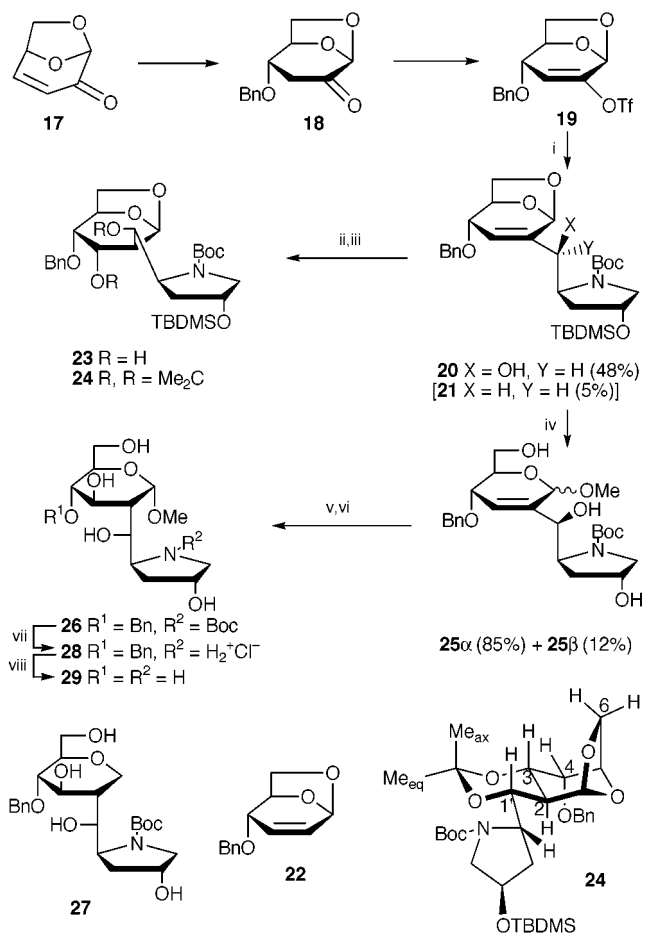
Hydroboration of **9** with BH<sub>3</sub>·SMe<sub>2</sub> in THF at 50 °C followed by H<sub>2</sub>O<sub>2</sub>/NaOH work-up furnished the D-glucose derivative **12** in 57% yield. Desilylation with Bu<sub>4</sub>NF in THF gave triol **13** (97%), which was debenzylated to give tetrol **14** (94%). Treatment of **14** in refluxing MeOH saturated with gaseous HCl for 2 days produced a 2:1 mixture of **15α** and **15β** in 83% yield.

The D-*gluco* configuration of the methyl pyranosides **15α** and **15β** was determined from their <sup>1</sup>H NMR spectra.<sup>13</sup> The configuration of the hydroxymethano linker was established by converting diol **12** into the corresponding acetonide **16** [(MeO)<sub>2</sub>CMe<sub>2</sub>, acetone, TsOH, Drierite, 25 °C, 68% yield], the structure of which was established by its NOESY and <sup>1</sup>H NMR spectra. The coupling constants, <sup>3</sup>J(H-3, H-4) = 10.9 Hz, <sup>3</sup>J(H-4, H-1') = 11.2 Hz and <sup>3</sup>J(H-2, H-3) = 7.2 Hz, show the antiperiplanar orientation between these proton pairs. Furthermore, NOEs were observed between signals attributed to H<sub>syn</sub>-6, H-2, H-4, and between those attributed to Me<sub>axial</sub>, H-1', H-3. Thus, the *Re* face of aldehyde **8** is preferred for the alkenylchromium addition, which is in accord with the Felkin–Anh Model.<sup>14</sup>



**Scheme 2** Reagents and conditions: i, BnOH, Et<sub>3</sub>N, 25 °C; ii, LiHMDS, HMPA–THF, 2-[bis(trifluoromethylsulfonyl)amino]-5-chloropyridine, –78 °C, 87%; iii, **8**, CrCl<sub>2</sub>, NiCl<sub>2</sub>, O<sub>2</sub>, DMF, ultrasound, 25 °C; iv, BH<sub>3</sub>·SMe<sub>2</sub>, THF, 50 °C, v, H<sub>2</sub>O<sub>2</sub>, NaOH, 57% for 2 steps; vi, Bu<sub>4</sub>NF, THF, 97%; vii, H<sub>2</sub>, 10% Pd–C, MeOH, 25 °C, 94%; viii, MeOH, HCl, reflux, 2 days, 83%.

† Spectral data for **15α**, **15β** and **29** are available from the RSC web site, see <http://www.rsc.org/suppdata/cc/1999/1873/>



**Scheme 3** Reagents and conditions: i, **8**, CrCl<sub>2</sub>, NiCl<sub>2</sub>, O<sub>2</sub>, DMF, ultrasound, 25 °C; ii, BH<sub>3</sub>·SMe<sub>2</sub>, THF, reflux; iii, H<sub>2</sub>O<sub>2</sub>, NaOH, 37% for 2 steps; iv, MeOH, TsOH, 25 °C; v, BH<sub>3</sub>·SMe<sub>2</sub>, THF; vi, H<sub>2</sub>O<sub>2</sub>, NaOH, 46% for 2 steps; vii, MeOH, HCl, reflux, 92%; viii, H<sub>2</sub>, 10% Pd-C, MeOH, 25 °C, 95%.

In parallel with the synthesis of (1→4)-C-disaccharides, (1→2)-C-disaccharides and analogues can be obtained starting from levoglucosenone **17** (Scheme 3). Benzyl alcohol adduct **18** was converted (as above) into triflate **19** in 90% yield.<sup>15</sup> In the Nozaki-Kishi coupling reaction, **19** was found to be less reactive than triflate **7**. It required activation with ultrasound and a catalytic amount of O<sub>2</sub> to react (best results with 5 mol% O<sub>2</sub> with respect to CrCl<sub>2</sub>, 0.3 mol% NiCl<sub>2</sub>). This led to alcohol **20** isolated in 48% yield, together with dehydroxylated product **21** (5% yield) and side product **22** (8% yield).<sup>‡</sup> Hydroboration of **20** followed by oxidative work-up provided diol **23** in modest yield (37%). The <sup>1</sup>H NMR and NOESY spectra of acetone **24** (obtained under the same conditions as those for **16**) showed coupling constants <sup>3</sup>J(H-2, H-3) = 10.7 Hz, <sup>3</sup>J(H-1', H-2) = 10.7 Hz, <sup>3</sup>J(H-3, H-4) = 3.6 Hz, and NOEs between proton pairs H<sub>syn</sub>-6/H-3, H-3/H-1' H-3/Me<sub>axial</sub> and H-1'/Me<sub>axial</sub>. These establish the *D-altra* configuration of the anhydrohexose moiety and the (*R*) configuration of the hydroxymethano linker demonstrating again that the *Re* face of **8** was preferred for the nucleophilic addition.<sup>16</sup> Hydroboration took place from the *exo* face of the bicyclic system probably because of steric hindrance from the *endo* hydrogen at C6. The allylic acetal was readily opened by acidic methanolysis, which provided glycosides **25α** and **25β** isolated in 85 and 12% yield, respectively. Hydroboration of **25α** gave alcohol **26** (46%) and the anhydroglucitol derivative **27** (10%).<sup>16</sup> Under reflux in MeOH saturated with HCl, **28** was obtained in 92% yield. Hydrogenation liberated the (1→2)-C-linked imino disaccharide **29** (95%).

The stereoselective methods presented above should be applicable to the preparation of a large variety of (1→2)- and (1→4)-C-disaccharides and analogues employing the same starting materials as those for the synthesis of (1→3)-C-

disaccharides, and thus make possible the exploration of their structure-activity relationships.<sup>17</sup>

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## Notes and references

<sup>‡</sup> Selected data for **9**: colorless oil; [α]<sub>D</sub><sup>25</sup> -107 (c 1.2, CHCl<sub>3</sub>); δ<sub>H</sub>(400 MHz, CDCl<sub>3</sub>, 323 K) 7.38–7.25 (m, 5H, Ph), 5.54 (m, 2H, H-1, H-3), 4.76 [d, <sup>3</sup>J(H-5, H<sub>exo</sub>-6) 3.6, H-5], 4.65 and 4.62 (2d, 2H, <sup>2</sup>J 11.5, PhCH<sub>2</sub>O), 4.38 (p, <sup>3</sup>J 3.6, H-4'), 4.37 (m, H-1'), 4.31 (m, H-2'), 3.71 (m, 2H, H-6), 3.55 [d, <sup>3</sup>J(H-2, H-3) 3.9, H-2), 3.48 (d, <sup>2</sup>J 11.2, H-5'a), 3.35 [dd, <sup>2</sup>J 11.2, <sup>3</sup>J(H-4', H-5'b) 4.2, H-5'b] 1.92 (m, 2H, H-3'), 1.48 [s, 9H, (CH<sub>3</sub>)<sub>3</sub>CO], 0.86 [s, 9H, (CH<sub>3</sub>)<sub>3</sub>CSi], 0.04 [s, 6H, (CH<sub>3</sub>)<sub>3</sub>Si]. For **20**: [α]<sub>D</sub><sup>25</sup> +24 (c 0.94, CHCl<sub>3</sub>); δ<sub>H</sub>(400 MHz, CDCl<sub>3</sub>, 323 K) 7.44–7.26 (m, 5H, Ph), 5.69 (m, H-3), 5.64 (s, H-1), 4.77 [dtd, <sup>3</sup>J(H-5, H<sub>exo</sub>-6) 6.6, <sup>4</sup>J(H-3, H-5) = <sup>3</sup>J(H-5, H<sub>endo</sub>-6) = 1.7, <sup>3</sup>J(H-4, H-5) 1.1, H-5), 4.68 (s, 2H, PhCH<sub>2</sub>O), 4.52 (m, H-1'), 4.32 (m, H-4'), 4.20 (m, H-2'), 3.87 (dd, <sup>2</sup>J 7.6, <sup>3</sup>J 6.6, H<sub>exo</sub>-6), 3.58 [dt, <sup>3</sup>J(H-3, H-4) 3.9, <sup>3</sup>J(H-4, H-5) = <sup>5</sup>J(H-4, H-1') = 1.1, H-4], 3.46 (m, H-5'a), 3.36 (dd, <sup>2</sup>J 11.2, <sup>3</sup>J 4.2, H-5'b), 3.31 (dd, <sup>2</sup>J 7.6, <sup>3</sup>J 1.7, H<sub>endo</sub>-6), 1.89 (m, 2H, H-3'), 1.49 [s, 9H, (CH<sub>3</sub>)<sub>3</sub>CO], 0.88 [s, 9H, (CH<sub>3</sub>)<sub>3</sub>CSi], 0.06 [s, 6H, (CH<sub>3</sub>)<sub>2</sub>Si].

- See, e.g.: A. Varki, *Glycobiology*, 1993, **3**, 97; P. R. Crocker and T. Feizi, *Curr. Opin. Struct. Biol.*, 1996, **6**, 679; A. Varki, *Proc. Natl. Acad. Sci. U.S.A.*, 1994, **91**, 7390; K. W. Moremen, R. B. Trimble and A. Herscovics, *Glycobiology*, 1994, **4**, 113; B. Ganem, *Acc. Chem. Res.*, 1996, **29**, 340 and references cited therein.
- R. Kornfeld and S. Kornfeld, *Annu. Rev. Biochem.*, 1985, **54**, 631; A. Lai, P. Pang, S. Kalelkar, P. A. Romero, A. Herscovics and K. W. Moremen, *Glycobiology*, 1998, **8**, 981 and references cited therein.
- J. Marshall, *J. Adv. Carbohydr. Chem. Biochem.*, 1974, **30**, 257; D. D. Schmidt, W. Frommer, B. Junge, L. Müller, W. Wingender, E. Truscheit and D. Schefer, *Naturwissenschaften*, 1977, **64**, 535.
- A. D. Elbein and R. J. Molyneux, in *Iminosugars as Glycosidase Inhibitors*, ed. A. E. Stütz, Wiley-VCH, Weinheim, 1999, pp. 216–251; C. W. Eckhart, M. H. Fechter, P. Hadwiger, E. Mlaker, A. E. Stütz, A. Tauss and T. M. Wrodnigg, in *Iminosugars as Glycosidase Inhibitors*, ed. A. E. Stütz, Wiley-VCH, Weinheim, 1999, pp. 253–390; M. Bols, *Acc. Chem. Res.*, 1998, **31**, 1; E. Fenouillet, M. J. Papandreou and I. M. Jones, *Virology* 1997, **231**, 89; T. Kolter, *Angew. Chem., Int. Ed. Engl.*, 1997, **36**, 1955 and references cited therein.
- C. Pasquarello, R. Demange and P. Vogel, *Bioorg. Med. Chem. Lett.*, 1999, **9**, 793.
- B. A. Jones, Y. T. Pan, A. D. Elbein and C. R. Johnson, *J. Am. Chem. Soc.*, 1997, **119**, 4856; K. Kraehenbuehl, S. Picasso and P. Vogel, *Helv. Chim. Acta*, 1998, **81**, 1439; M. A. Leeuwenburgh, S. Picasso, H. S. Overkleef, G. A. van der Marel, P. Vogel and J. H. van Boom, *Eur. J. Org. Chem.*, 1999, 1185.
- See, e.g.: G. D. MacLean, M. B. Bowen-Yacyshyn, J. Samuel, A. Meikle, G. Stuart, J. Nation, S. Poppema, M. Jerry, R. Koganty, T. Wong and B. M. Longenecker, *J. Immunother.*, 1992, **11**, 292; P. D. Rye, N. V. Bovin, E. V. Vlasova and R. A. Walker, *Glycobiology*, 1995, **5**, 385.
- Y.-H. Zhu and P. Vogel, *Tetrahedron Lett.*, 1998, **39**, 31.
- Y.-H. Zhu and P. Vogel, *J. Org. Chem.*, 1999, **64**, 666.
- D. L. Comins and A. Dehghani, *Tetrahedron Lett.*, 1992, **33**, 6299.
- S. Mori, T. Ohno, H. Harada, T. Aoyama and T. Shioiri, *Tetrahedron*, 1991, **47**, 5051.
- K. Takai, M. Tagashira, T. Kuroda, K. Oshima, K. Uimoto and H. Nozaki, *J. Am. Chem. Soc.*, 1986, **108**, 6048; D. P. Stamos, X. C. Sheng, S. S. Chen and Y. Kishi, *Tetrahedron Lett.*, 1997, **38**, 6355 and references cited therein.
- Y. Wang, P. G. Goekjian, D. M. Ryckman, W. H. Miller, S. A. Babirad and Y. Kishi, *J. Org. Chem.*, 1992, **57**, 482.
- F. Rübsam, S. Seck and A. Giannis, *Tetrahedron*, 1997, **53**, 2823; P. Ciapetti, M. Falorni and M. Taddei, *Tetrahedron*, 1996, **52**, 7379.
- For the preparation of **18**, see: T. Kawai, M. Isobe and S. C. Peters, *Aust. J. Chem.*, 1995, **48**, 115.
- J. Cossy, V. Bellosta and M. C. Müller, *Tetrahedron Lett.*, 1992, **33**, 5045.
- An example of the synthesis of the β-C(1→4)glucopyranoside of 3-deoxy-D-glucose from levoglucosenone was reported by Witczak and co-workers: Z. J. Witczak, R. Chhabra and J. Chojnacki, *Tetrahedron Lett.*, 1997, **38**, 2215.